MBPCR026 Campylobacter jejuni Detection Kit (Real-Time)

Description:

Campylobacter jejuni is a Gram-negative slender, curved, motile rod shaped, micro-aerophilic organism which requires less oxygen to survive. It is relatively fragile and sensitive to environmental stresses. This bacterium is recognized as an important enteric pathogen (intestinal bacteria). It mainly spreads through raw and undercooked poultry, raw milk and untreated water. *C. jejuni* frequently contaminates raw chicken.

Campylobacteriosis, often known as campylobacter enteritis or gastroenteritis, is the name of the infection caused by *C. jejuni*. *C. jejuni* infection causes diarrhea and fecal leukocytes (white cells). Other symptoms often present are fever, abdominal pain, nausea, headache and muscle pain. The pathogenic mechanisms of *C. jejuni* are still not completely understood, but it does produce a heat-labile toxin that may cause diarrhea.

Specific and faster methods for detection of foodborne pathogens, such as real–time PCR, are the need of an hour. These techniques help to detect targeted pathogens quickly; this early and precise detection helps to take further actions.

NOTE: The Campylobacter jejuni Detection Kit (Real-Time) is for *in vitro* use only.

Principle:

The Campylobacter jejuni Detection Kit (Real-Time) is designed for detection of specific sequence of **MDmapA** gene giving amplification of **589 bp** product for *C. jejuni* in various food sources, cells, clinical samples etc. The kit allows rapid, sensitive and specific detection of *Campylobacter jejuni*.

Real-time Polymerase Chain Reaction, also called quantitative Polymerase Chain Reaction (qPCR) or kinetic Polymerase Chain Reaction, is a laboratory technique based on the principle of Polymerase Chain Reaction. This technique is used to amplify and simultaneously quantitate a targeted DNA sequence. Real-time PCR systems based on SYBr Green assays have increasingly been used for accurate, reliable detection and quantitation of various food-borne pathogens. HiMedia's Campylobacter jejuni Detection kit (Real-time), is one such SYBr green based qPCR techinique which allows amplification of **MDmapA gene.**

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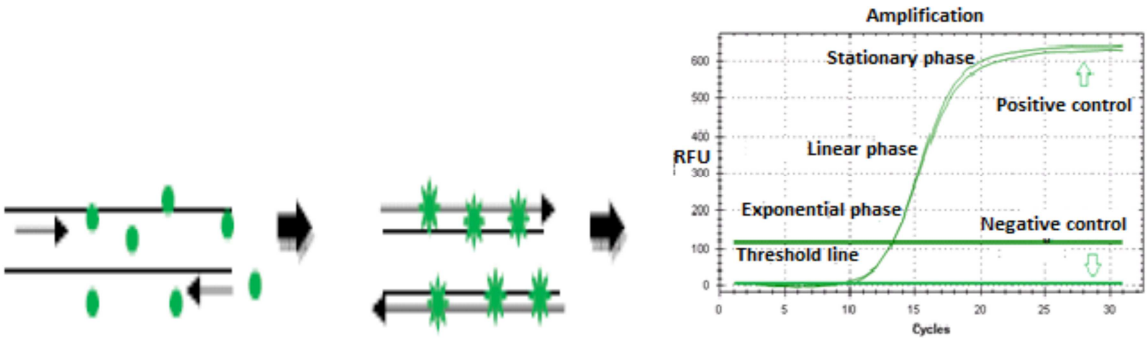
A)Diagrammatic representation of preferential binding of SYBr Green Dye to specific DNA fragments in real-time PCR.

a) Dye in solution emits low fluorescence

b) Emission of the fluorescence by binding

c) Amplification data



SYBr Green dye cycles between an unbound (Denaturation step) and a bound (Annealing through Extension) state as the reaction progresses. Signal intensity increases as the quantity of amplicons increase in later cycles indicating amplification. During elongation, more and more dye molecules bind to the newly synthesized DNA. If the reaction is monitored continuously, an increase in fluorescence is viewed in real-time. Upon denaturation of the DNA for the next heating cycle, the dye molecules are released and the fluorescence signal falls.

Keys : SYBr
Forward primer
Reverse primer
DNA Strand

Features:

- Fast and simple
- Good sensitivity and specific results
- Guaranteed reproducible results
- Rapid detection of all relevant clinical pathogens

Kit Contents:

The provided PCR kit contains:

Components	Reagents provided for 10R (reactions)*	Reagents provided for 25R (reactions)*	Reagents provided for 50R (reactions)*
Hi-SYBr master mix (2X master mix containing SYBr Green, Assay buffer, Taq Polymerase, $MgCl_2$, dNTPs) **(MBT074)**	150 µl	400 µl	700 µl
Primer Mix	25 µl	60 µl	120 µl
Nuclease free water **(ML065)**	1 ml	2 ml	4 ml

***** For a 20µl PCR reaction

General Preparation Instructions:

- Before use all PCR components should be completely thawed on ice (4°C).

- Perform the amplification reactions in a clean area, preferably in a biosafety cabinet.
- Use of aerosol barrier pipette tips is recommended to reduce contamination risks from extraneous DNA templates.
- Extract and store positive control sample (if used) separately from all other reagents to avoid contamination and add it to the reaction mix in a separate area.

Sampling and Handling:

Sample Preparation:

Various food, clinical and environmental samples as well as cultured bacteria are routinely examined.

For extraction and purification of high yield and pure bacterial DNA, perform the nucleic acid purification using HiMedia's **HiPurA™ Bacterial Genomic DNA Purification Kit (MB505)** as instructed in the protocol.

Flow Chart for setting up PCR Reaction

Add 10µl Hi-SYBr master mix **(MBT074)** in a PCR tube

⇩

Add 2 µl the Primer mix (Final concentration 10 pmoles provided)

⇩

Add 1-2 µl template DNA (upto 50 ng of extracted DNA)

⇩

Add nuclease free water (ML065) to make the final volume to 20 µl

⇩

Centrifuge the tube briefly at 6000 rpm for about 10 seconds.

⇩

Place the tubes in real-time PCR machine and set the recommended PCR program (mentioned below)

⇩

Interpret the data from the amplification plot (observe the Ct values)

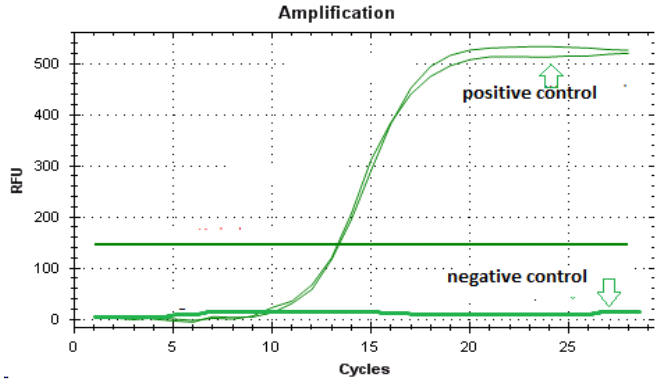
Recommended PCR program:

- Initial denaturation : 95°C for 10 minutes

- Cycling Parameters (No. of cycles: 30)
  Denaturation : 95°C for 30 seconds
  Annealing : 59°C for 90 seconds
  Extension : 72°C for 60 seconds

- Final Extension : 72°C for 10 minutes.

Amplification Data:



Sr. No.	Sample	C_t value
1	Negative control	N/A
2	1 µl of template DNA (amplicon of *C. jejuni*	13.40
3	1 µl of template (amplicon of *C. jejuni*)	13.48

Figure: Data representing real-time amplification data of *C. jejuni* with C_t values (provided in table)

Sensitivity: Detectable upto 100-1000 CFU / ml (mg).

Storage:

The provided kit has a shelf-life of 6 months when stored at -20˚C. Repeated thawing and freezing of PCR reagents should be avoided, as this may reduce the sensitivity. If reagents are to be used multiple times, we recommend storing reagents as aliquots to avoid repeated freeze and thaw. Degradation of sample DNA specimens can also reduce sensitivity of the assay. HiMedia does not recommend using the kit after the expiry date stated on pack.

Quality Control:

Each lot of HiMedia's Campylobacter jejuni Detection Kit (Real-time) is assayed for contaminating endonuclease, exonuclease and non-specific DNase activities. Functionally tested in DNA amplification.

Troubleshooting Guide:

Sr.No.	Problem	Cause	Solution
1.	No amplification	Degraded samples	1. Check the integrity of DNA using agarose gel electrophoresis.
			2. Use freshly prepared DNA to ensure the availability of intact template sequence for efficient amplification.
		Error in protocol setup	Verify that the correct reagent volumes, dilutions and storage conditions have been used.
2.	Variability between replicates	Error in reaction set-up	Prepare large volume master mix, vortex thoroughly and aliquot into reaction tubes.
		Air bubbles in reaction mix	Briefly centrifuge reaction samples/plate prior to running on a real-time PCR instrument.
		Pipetting error	C_t values of replicates can show increased variation due to poor laboratory technique or imprecise pipettes.

3.	Amplification in negative control	Reagents contaminated	1. Replace all critical solutions 2. Repeat the analysis of all tests with fresh aliquots of critical reagents.

Safety Information

The Campylobacter jejuni Detection Kit (Real-time) is for laboratory use only, not for drug, household or other uses. Take appropriate laboratory safety measures and wear gloves when handling.

Product Use Limitation & Warranty

HiMedia guarantees the performance of Campylobacter jejuni Detection Kit (Real-time) in the manner described in the product literature. The kit is designed, sold for research and for *in vitro* purposes only. No claim or representation is intended to provide information for the diagnosis, prevention or treatment of a disease.
All due care and attention should be exercised in the handling of the products. We recommend all users of HiMedia products to adhere to the NIH guidelines that have been developed for recombinant DNA experiments, or to other applicable guidelines.

Technical Assistance

At HiMedia, we pride ourselves on the quality and availability of our technical support. For any kind of technical assistance, mail at mb@himedialabs.com.

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